



Lot

Page 1 of 2

Full activity is guaranteed through the date indicated on the Certificate of Analysis.

Catalog Number	200152
Product Name	SURE® 2 Supercompetent Cells
Materials Provided	SURE® 2 supercompetent cells (red-orange tubes), 10 × 100 µl pUC18 control plasmid (0.1 ng/µl in TE buffer), 10 µl β-Mercaptoethanol (1.22 M), 25 µl
Certified By	****
Quality Controlled By	****
Shipping Conditions	Shipped on dry ice.
Storage Conditions	Competent cells must be placed immediately at the bottom of a -80°C freezer directly from the dry ice shipping container. Do not store the cells in liquid nitrogen. Competent cells are sensitive to even small variations in temperature. Transferring tubes from one freezer to another may result in a loss of efficiency.
Guaranteed Efficiency	≥1.0 × 10 ⁹ cfu/µg pUC18 DNA
Test Conditions	Transformations are performed both with and without plasmid DNA using 100-µl aliquots of cells and 10 pg of pUC18 control DNA following the protocol outlined below. Following transformation, 5-µl samples of the culture are plated in duplicate on LB agar plates with 100 µg/ml ampicillin. The plates are incubated at 37°C overnight and the efficiency is calculated based on the average number of colonies per plate.
Genotype and Background	<p>e14(McrA⁻) Δ(<i>mcrCB-hsdSMR-mrr</i>)171 <i>endA1 gyrA96 thi-1 supE44 relA1 lac recB recJ sbcC umuC::Tn5</i> (Kan^r) <i>uvrC</i> [F' <i>proAB lacI^qΔM15 Tn10</i> (Tet^r) Amy Cam^r]. (Genes listed signify mutant alleles. Genes on the F' episome, however, are wild-type unless indicated otherwise).</p> <p>SURE® 2 supercompetent cells* are high-efficiency derivatives of Stratagene's SURE (Stop Unwanted Rearrangement Events) competent cells, which have been engineered to allow the cloning of certain DNA segments that are "unclonable" in conventional <i>E. coli</i> strains. The SURE strain lacks components of the pathways that catalyze the rearrangement and deletion of nonstandard secondary and tertiary structures, including cruciforms (caused by inverted repeats) and Z-DNA, that occur frequently in eukaryotic DNA and that impede the cloning of the eukaryotic DNA in conventional strains. SURE 2 cells are restriction minus (McrA⁻, McrCB⁻, McrF⁻, Mrr⁻, HsdR⁻) endonuclease (<i>endA</i>) deficient, and recombination (<i>recB recJ</i>) deficient. The <i>lacI^qΔM15</i> gene, on the F' episome, allows blue-white screening.</p>
Antibiotic Resistance	SURE® 2 cells are kanamycin, tetracycline, and chloramphenicol resistant.
Transformation Protocol	<ol style="list-style-type: none">1. Pre-chill two 14-ml BD Falcon polypropylene round-bottom tubes on ice. (One tube is for the experimental transformation and one tube is for the pUC18 control.) Preheat NZY⁺ medium to 42°C.2. Thaw the cells on ice. When thawed, gently mix and aliquot 100 µl of cells into each of the two pre-chilled tubes.3. Add 2 µl of the β-ME provided with this kit to each aliquot of cells.4. Swirl the tubes gently. Incubate the cells on ice for 10 minutes, swirling gently every 2 minutes.5. Add 0.1-50 ng of the experimental DNA to one aliquot of cells, in a 1-µl volume for maximum efficiency. Dilute the pUC18 control DNA 1:10 with sterile dH₂O, then add 1 µl of the diluted pUC18 DNA to the other aliquot of cells.6. Swirl the tubes gently, then incubate the tubes on ice for 30 minutes.7. Heat-pulse the tubes in a 42°C water bath for 30 seconds. The duration of the heat pulse is critical.8. Incubate the tubes on ice for 2 minutes.9. Add 0.9 ml of preheated (42°C) NZY⁺ broth and incubate the tubes at 37°C for 1 hour with shaking at 225-250 rpm.10. Plate ≤200 µl of the transformation mixture on LB agar plates containing the appropriate antibiotic (and containing IPTG and X-gal if color screening is desired). For the pUC18 control transformation, plate 5 µl of the transformation mixture on LB-ampicillin agar plates.11. Incubate the plates at 37°C overnight. If performing blue-white color screening, incubate the plates at 37°C for at least 17 hours to allow color development (color can be enhanced by subsequent incubation of the plates for 2 hours at 4°C).12. For the pUC18 control, expect 50 colonies (≥1 × 10⁹ cfu/µg pUC18 DNA). For the experimental DNA, the number of colonies will vary according to the size and form of the transforming DNA, with larger and non-supercoiled DNA producing fewer colonies.

Blue-White Color Screening

Blue-white color screening for recombinant plasmids is available when transforming this host strain (containing the *lacI^qΔM15* gene on the F' episome) with a plasmid that provides α-complementation (e.g. Stratagene's pBluescript[®]II vector). When *lacZ* expression is induced by IPTG in the presence of the chromogenic substrate X-gal, colonies containing plasmids with inserts will be white, while colonies containing plasmids without inserts will be blue. If an insert is suspected to be toxic, plate the cells on media without X-gal and IPTG. Color screening will be eliminated, but lower levels of the potentially toxic protein will be expressed in the absence of IPTG.

Critical Success Factors and Troubleshooting

Use of 14-ml BD Falcon polypropylene round-bottom tubes: It is important that 14-ml BD Falcon polypropylene round-bottom tubes (BD Biosciences Catalog #352059) are used for the transformation protocol, since other tubes may be degraded by β -mercaptoethanol. In addition, the duration of the heat pulse has been optimized using these tubes.

Aliquoting Cells: Keep the cells on ice at all times during aliquoting. It is essential that the polypropylene tubes are placed on ice before the cells are thawed and that the cells are aliquoted directly into pre-chilled tubes. It is also important to use 100 μ l of cells per transformation. Decreasing the volume will reduce efficiency.

Use of β -Mercaptoethanol (β -ME): β -ME has been shown to increase transformation efficiency. The β -ME mixture provided is diluted and ready to use. Stratagene cannot guarantee results with β -ME from other sources.

Use of NZY⁺ Broth: Transformation of the supplied supercompetent cells has been optimized using NZY⁺ as the medium for outgrowth following the heat pulse. Substitution with another outgrowth medium may result in a loss of efficiency.

Quantity and Volume of DNA: The greatest efficiency is obtained from the transformation of 1 μ l of 0.01 ng/ μ l supercoiled pUC18 DNA per 100 μ l of cells. When transforming a ligation mixture, add 2 μ l of the ligation mixture per 100 μ l of cells. A greater number of colonies may be obtained by transforming up to 50 ng DNA, although the resulting efficiency (cfu/ μ g) may be lower. The volume of the DNA solution added to the reaction may be increased to up to 10% of the reaction volume, but the transformation efficiency may be reduced.

Heat Pulse Duration and Temperature: Optimal transformation efficiency is observed when cells are heat-pulsed at 42°C for 30 seconds. Efficiency decreases sharply when cells are heat-pulsed for <30 seconds or for >40 seconds. Do not exceed 42°C.

Plating the Transformation Mixture: If plating <100 μ l of cells, pipet the cells into a 200- μ l pool of NZY⁺ medium and then spread the mixture with a sterile spreader. If plating \geq 100 μ l, the cells can be spread on the plates directly. Tilt and tap the spreader to remove the last drop of cells. If desired, cells may be concentrated prior to plating by centrifugation at 1000 rpm for 10 minutes followed by resuspension in 200 μ l of NZY⁺ medium.

Preparation of Media and Reagents**NZY⁺ Broth (per Liter)**

10 g of NZ amine (casein hydrolysate)

5 g of yeast extract

5 g of NaCl

Add deionized H₂O to a final volume of 1 liter

Adjust to pH 7.5 using NaOH and then autoclave

Add the following filter-sterilized supplements prior to use:

12.5 ml of 1 M MgCl₂

12.5 ml of 1 M MgSO₄

20 ml of 20% (w/v) glucose (or 10 ml of 2 M glucose)

LB Agar (per Liter)

10 g of NaCl

10 g of tryptone

5 g of yeast extract

20 g of agar

Add deionized H₂O to a final volume of 1 liter

Adjust pH to 7.0 with 5 N NaOH and then autoclave

Pour into petri dishes (~25 ml/100-mm plate)

LB-Ampicillin Agar (per Liter)

1 liter of LB agar, autoclaved and cooled to 55°C

Add 10 ml of 10 mg/ml filter-sterilized ampicillin

Pour into petri dishes (~25 ml/100-mm plate)

Plates for Blue-White Color Screening

Prepare the LB agar and when adding the antibiotic, also add 5-bromo-4-chloro-3-indolyl- β -D-galactopyranoside (X-gal) to a final concentration of 80 μ g/ml [prepared in dimethylformamide (DMF)] and isopropyl- β -D-galactopyranoside (IPTG) to a final concentration of 20 mM (prepared in sterile water). Alternatively, 100 μ l of 10 mM IPTG and 100 μ l of 2% X-gal may be spread on solidified LB agar plates 30 minutes prior to plating the transformations. (For consistent color development across the plate, pipet the X-gal and the IPTG into a 100- μ l pool of SOC medium and then spread the mixture across the plate. Do not mix the IPTG and the X-gal before pipetting them into the pool of SOC medium because these chemicals may precipitate.)

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Endnotes

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