

Sensitive Femtogram Determination of Aflatoxins B₁, B₂, G₁, and G₂ in Food Matrices using Tandem LC-MS/MS

Yang Chen¹, Peter Stone² and Jack Cappozzo¹

¹National Center for Food Safety and Technology, Illinois Institute of Technology, IL

²Agilent Technologies Inc, Santa Clara, CA

Introduction

Aflatoxins are a group of mycotoxins produced as metabolites by the fungi *Aspergillus flavus* and *Aspergillus parasiticus* (1). They can be found in various foods including grains, nuts, and spices (2). There are four major naturally occurring aflatoxins: B₁, B₂, G₁, and G₂ (Fig 1). Exposure to them can cause cancer in humans and live stock. Reliable and sensitive analytical methods for the determination of aflatoxins are required to safeguard our food supply.

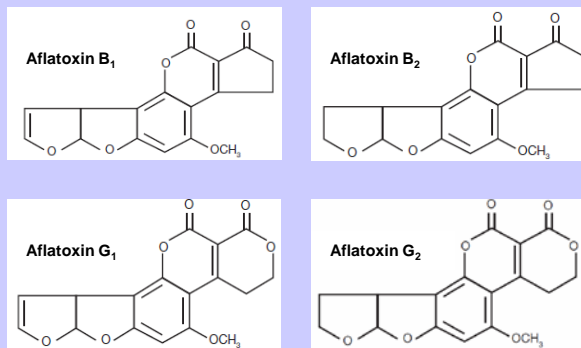


Fig 1. Structures of aflatoxins B₁, B₂, G₁ and G₂

Objectives

To develop a robust sample preparation and sensitive analytical method for the determination and confirmation of major aflatoxins in cereals and nuts.

Materials and Methods

LC/MS/MS Instrumentation

An Agilent 1200 series rapid resolution LC and a 6460 series Triple Quadrupole MS/MS system equipped with Agilent Jet Stream thermal focussing electrospray ion source was used (Fig 2). Aflatoxins were detected using electrospray ionization (ESI) in multiple reaction monitoring (MRM) mode. MassHunter Workstation software was used for identification and quantitation.

Aflatoxin standards and foods

Purified aflatoxin standards (B₁, B₂, G₁, and G₂) were obtained from Sigma-Aldrich. Aflatoxin-free corn flour, wheat, peanut and walnut samples obtained from a local grocery store were used for recovery studies.



Fig 2. Agilent LC/MS/MS system

HPLC conditions

Column: Agilent Zorbax Eclipse C18, 4.6 x 50 mm, 1.8 micron
Flow rate: 0.6 mL/min
Column temp: 40°C
Injection volume: 5 µL
Mobile phase: A: 10 mM ammonium acetate in water, B: methanol
Gradient:
Time (min) A (%) B (%)
0 95 5
5 0 100
6 0 100

MS conditions

Drying gas temp: 325°C Sheath gas temp: 350°C
Gas flow rate: 10 L/min Sheath gas flow: 11 L/min
Nebulizer pressure: 50 psi EMV: 400 V
Capillary voltage: 4000 V

Optimization of ionization and data acquisition parameters

The fragmentor voltage and collision energy for the MS/MS analysis were optimized for more specific and sensitive detection of aflatoxins (Table 1).

Table 1. Data acquisition parameters of MRM transitions for each aflatoxin

Aflatoxin	Retention time (min)	Fragmentor voltage (V)	Molecular weight	Precursor ion (m/z)	Product ions (m/z)	Collision energy (V)
B ₁	4.68	130	312.1	313.1	285.1	20
					241.0	35
					269.1	25
B ₂	4.57	130	314.1	315.1	287.1	25
					259.1	25
					243.0	40
G ₁	4.40	130	328.1	329.1	243.1	25
					311.1	20
					283.0	20
G ₂	4.26	130	330.1	331.1	313.1	25
					245.1	30
					285.1	25

Sample preparation and recovery studies

- Corn flour, ground wheat, peanut and walnut samples (10 g each) spiked with a mixture of 4 aflatoxin standards, each at 5 and 25 ppb.
- Extracted using 40 mL acetonitrile-water (84:16, v/v) for 30 min with shaking at room temperature.
- Extract clean-up using either C₁₈ powdered adsorbent material (ODS SPE bulk sorbent, Agilent) or MycoSep 226 multi-functional column (Romer).
- Aliquots (0.4 mL) of the cleaned up extracts were diluted with 0.6 mL 10 mM ammonium acetate in water.
- The sample was then centrifuged at 14,000 x g for 3 min prior to LC-MS/MS analysis.
- Each food matrix and spike level was conducted in 7 replicates

Results and Discussion

The use of precursor and product ions in MS analysis allowed for sensitive detection and confirmation of all four aflatoxins (Fig 3). The standard curves for all 4 aflatoxins showed a good linearity from 0.1 to 100 ppb with R² greater 0.9999 (Fig 4 & 5). Recoveries were between 85-110% for each of the aflatoxins for all four spiked food matrices, with the MycoSep clean-up method slightly better than the C₁₈ one for walnut samples (Fig 6). A limit of detection was determined to be <0.15 µg/kg and limit of quantitation <0.5 µg/kg for all four sample matrices.

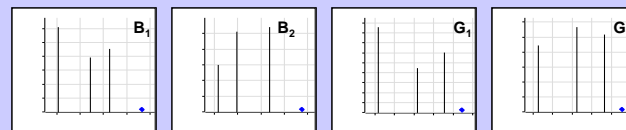


Fig 3. Mass spectra of aflatoxins B₁, B₂, G₁, and G₂.

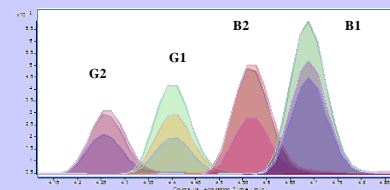


Fig 4. LC/MS/MS chromatogram of aflatoxin B₁, B₂, G₁, and G₂ standards at 1 ppb

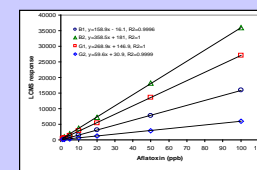


Fig 5. Standard curve of aflatoxins from 0.1 to 100 ppb

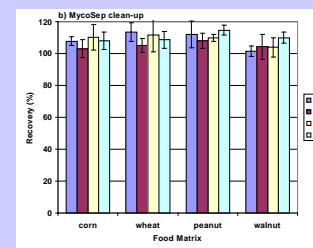
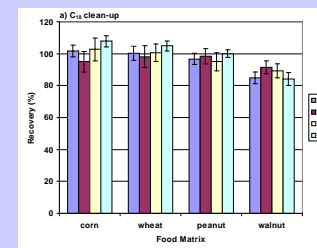


Fig 6. Recovery of aflatoxin B₁, B₂, G₁, and G₂ from food matrices using a) C₁₈ or b) MycoSep clean-up

Conclusions

An LC/MS/MS method has been developed for the analysis and confirmation of aflatoxins B₁, B₂, G₁, and G₂ in cereals and nuts, with a detection limit of less than 1 ppb.

References

- 1) Microchim Acts 153, 2006, 101-108
- 2) Rapid Commun. Mass Spectrum, 2009; 23: 3-11