Metabolic discovery platform: "Seahorse-Bravo"

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Metabolic Science for health

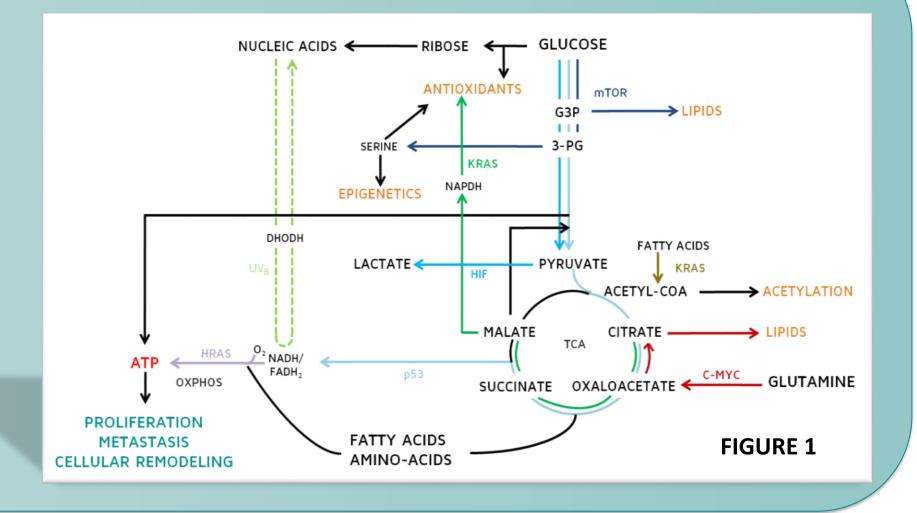
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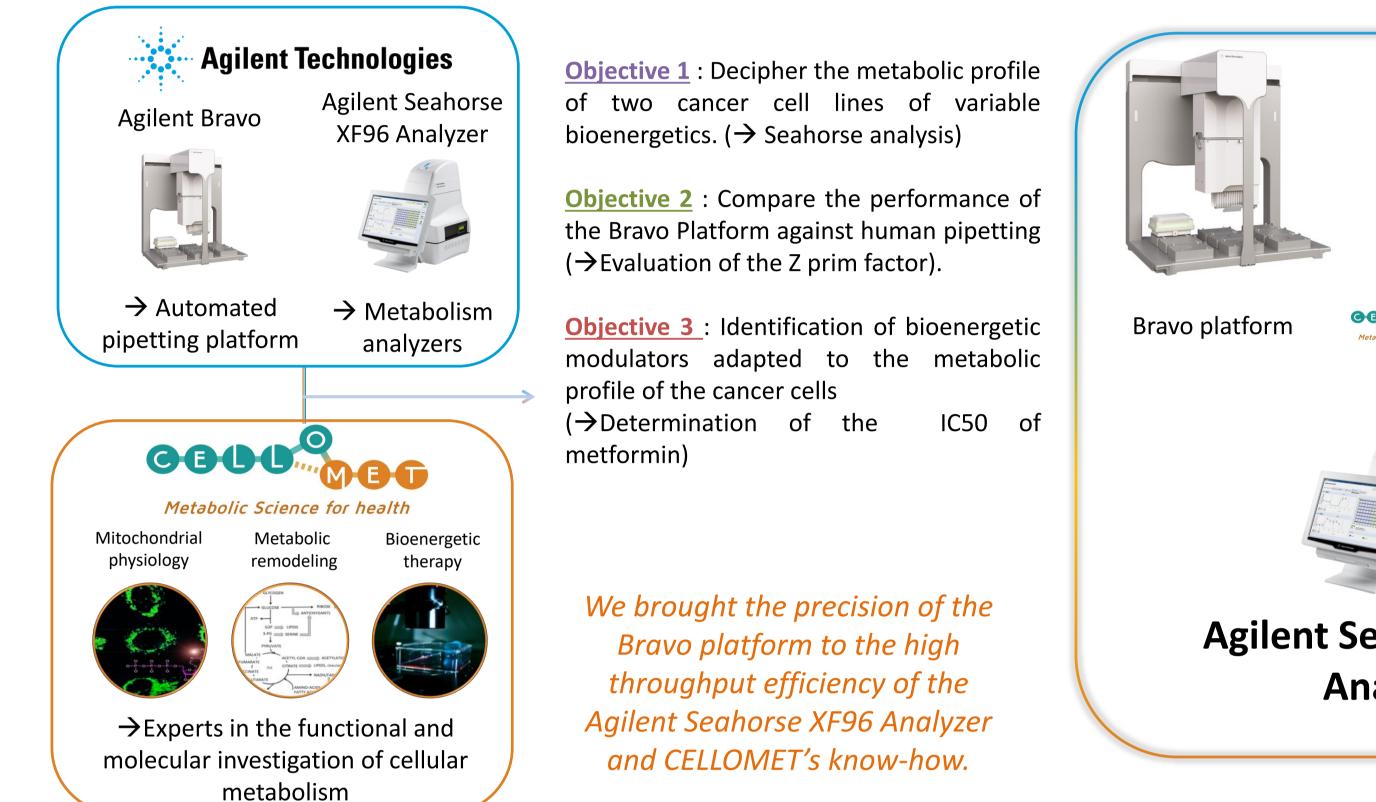
INTRODUCTION

Agilent Technologies

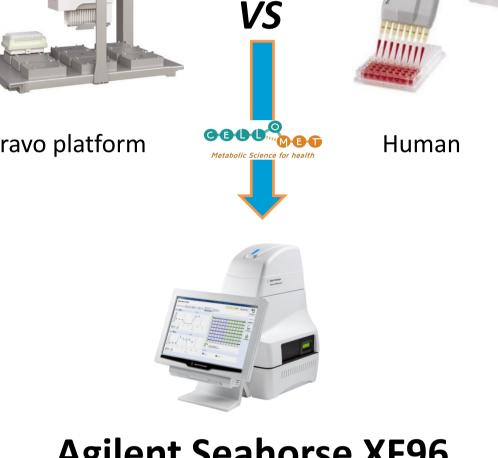
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The regulation of energy metabolism is very complex and involves a large number of signaling pathways and molecular effectors. Recent development in biology revealed that cancer cells can rewire most metabolic pathways, revealing novel means of bioenergetic regulation (Figure 1). For instance, it was shown that C-MYC oncogene controls glutamine-supported cellular respiration and lipid synthesis or that glycolysis bottlenecking by PKM2 promotes de novo serine biosynthesis. With the recently ascertained role of mitochondria in cancer biology, the search for novel regulators of mitochondrial respiration was performed using genetic means, as CRISPR-CAS9 or shRNA libraries, or pharmacological means such as drug-libraries. There is a growing need in academia and industry to dispose technology solutions enabling the discovery of mitochondrial respiration regulators (genes or compounds) using standardized and reproducible high-precision methods. The aim of this unprecedented CELLOMET-Agilent collaboration is to evaluate the quality and the efficiency of coupling the Agilent Seahorse XF96 Analyzer extracellular flux analyzer with the Agilent Bravo Automated Liquid Handling Platform.





For each objective we seeded 30.000 cells of each of the two lung cancer cell lines (A549 and H460) in each well of a 96-well plate. Respiration was measured using the Agilent Seahorse XF96 Analyzer at the CELLOMET bioenergetic investigation in Bordeaux (France). A mitostress kit was used in the cartridge plate to measure basal, minimal and uncoupled respiration rate.

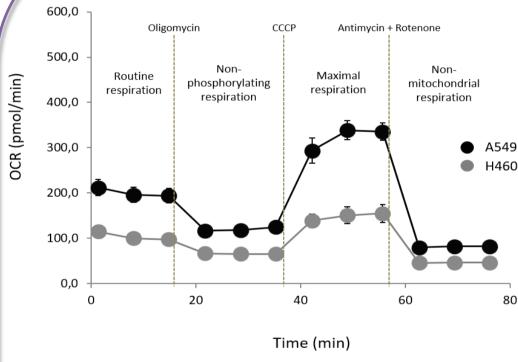


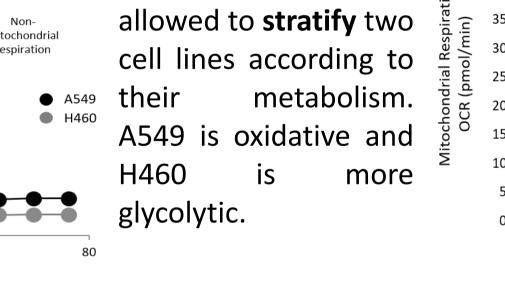
Agilent Seahorse XF96 Analyzer

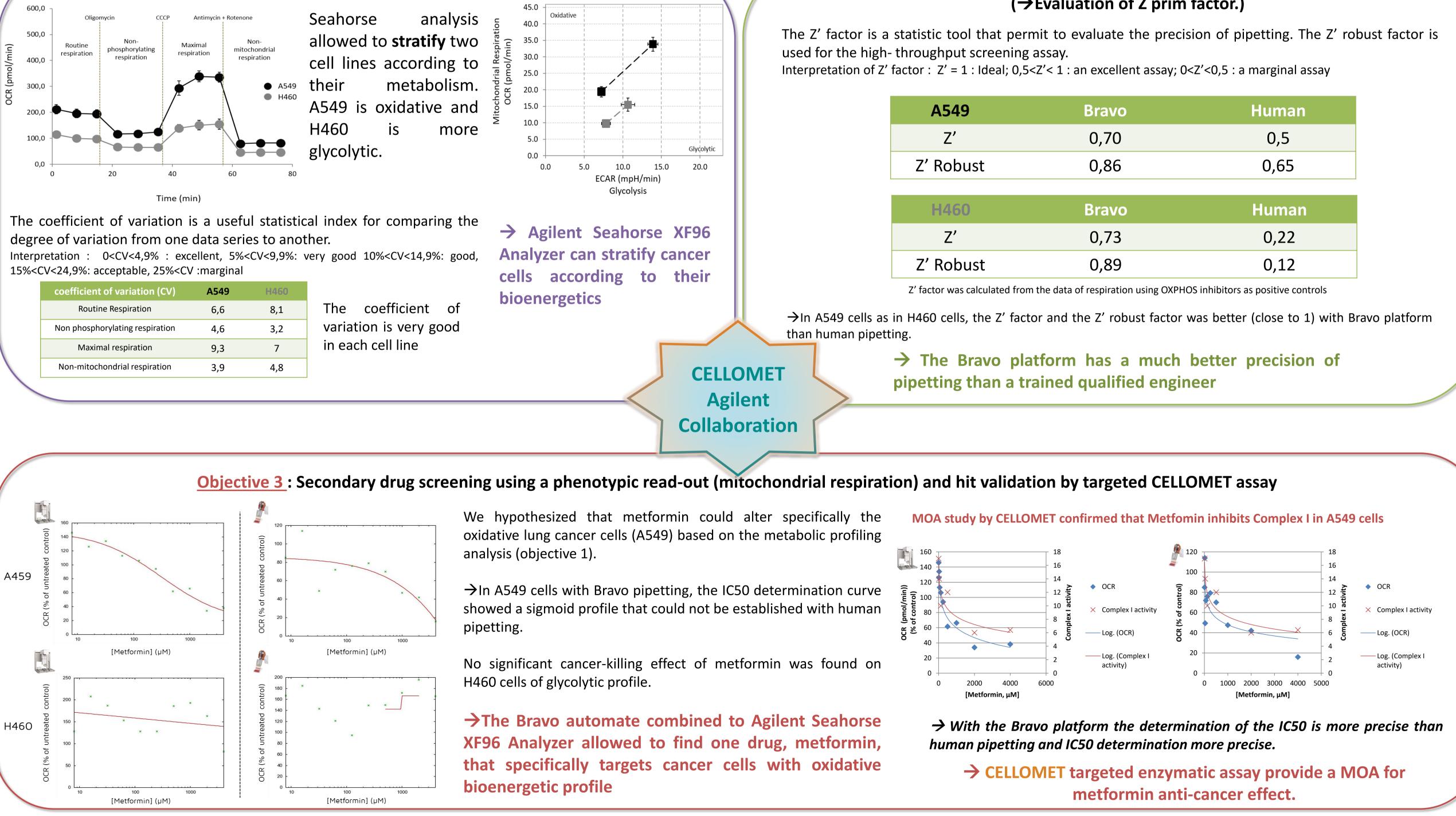
Z prim evaluation : The precision and reproducibility of our procedures can be evaluated by assessing the Z prim factor, which is a measure used for the GO/NO GO decision when performing high throughput screens of drugs or siRNA libraries. When precised, the cells were incubated with a cocktail of mitochondrial inhibitors for two hours. After this incubation time, a Seahorse test was performed at the CELLOMET facility.

IC50 of metformin test : The performance of the Bravo automate in the dilution of compounds was tested by the evaluation of the IC50 of metformin, a complex I inhibitor approved in the clinics for the treatment of diabetes. Metformin efficacy was tested on the cell lines of varying bioenergetics in cotreatment with doxorubicin, a valid chemotherapeutic drug (1µM). Different concentrations of metformin (0µM to 4mM) were tested, during 24h. To complete the results obtained on IC50 measurement with the Agilent Seahorse XF96 Analyzer, we performed a measure of respiratory chain complex I spectrophotometric activity using CELLOMET standardized protocol.

Objective 1: Bioenergetic profiling of two lung cancer cell lines







Objective 2 : Bravo platform versus human pipetting: Where's the difference? $(\rightarrow$ Evaluation of Z prim factor.)

A549	Bravo	Human
Z'	0,70	0,5
Z' Robust	0,86	0,65

H460	Bravo	Human
Ζ'	0,73	0,22
Z' Robust	0,89	0,12

CONCLUSION

Combining the Bravo Automated Liquid Handling Platform and the Agilent Seahorse XF96 Analyzer provides an accurate bioenergetic platform with excellent characteristics in terms of reproducibility and efficiency. Such platform can be used to study cellular energetics in a large number of contexts and to discover drug or gene modulations that impact mitochondrial respiration and cellular glycolysis. The Seahorse-Bravo equipment was validated by CELLOMET experts and a proof-of-concept experiment presented here showed the differences between A549 and H460 cells as well as their sensitivity to mitochondrial











